RESEARCH ARTICLE

Open Access



Essential oils from two *Allium* species exert effects on cell proliferation and neuroblast differentiation in the mouse dentate gyrus by modulating brain-derived neurotrophic factor and acetylcholinesterase

Hyo Young Jung¹, Kwon Young Lee², Dae Young Yoo¹, Jong Whi Kim¹, Miyoung Yoo³, Sanghee Lee³, Ki-Yeon Yoo⁴, Yeo Sung Yoon¹, Jung Hoon Choi^{2*} and In Koo Hwang^{1*}

Abstract

Background: In the present study, we investigated the effects of oil products from two *Allium* species: *Allium* sativum (garlic) and *Allium hookeri* (Chinese chives) on cell proliferation and neuroblast differentiation in the mouse dentate gyrus.

Methods: Using corn oil as a vehicle, the essential oil from garlic (10 ml/kg), or Chinese chives (10 ml/kg) was administered orally to 9-week-old mice once a day for 3 weeks. One hour following the last treatment, a novel object recognition test was conducted and the animals were killed 2 h after the test.

Results: In comparison to the vehicle-treated group, garlic essential oil (GO) treatment resulted in significantly increased exploration time and discrimination index during the novel object recognition test, while Chinese chives essential oil (CO) reduced the exploration time and discrimination index in the same test. In addition, the number of Ki67-immunoreactive proliferating cells and doublecortin-immunoreactive neuroblasts significantly increased in the dentate gyrus of GO-treated animals. However, administration of CO significantly decreased cell proliferation and neuroblast differentiation. Administration of GO significantly increased brain-derived neurotrophic factor (BDNF) levels and decreased acetylcholinesterase (AChE) activity in the hippocampal homogenates. In contrast, administration of CO decreased BDNF protein levels and had no significant effect on AChE activity, compared to that in the vehicle-treated group.

Conclusions: These results suggest that GO significantly improves novel object recognition as well as increases cell proliferation and neuroblast differentiation, by modulating hippocampal BDNF protein levels and AChE activity, while CO impairs novel object recognition and decreases cell proliferation and neuroblast differentiation, by reducing BDNF protein levels in the hippocampus.

Keywords: Acetylcholinesterase, Brain-derived neurotrophic factor, Chive, Garlic, Neurogenesis

Full list of author information is available at the end of the article



^{*} Correspondence: jhchoi@kangwon.ac.kr; vetmed2@snu.ac.kr

²Department of Anatomy, College of Veterinary Medicine and Institute of Veterinary Science, Kangwon National University, Chuncheon 24341, South Korea

¹Department of Anatomy and Cell Biology, College of Veterinary Medicine, Research Institute for Veterinary Science, Seoul National University, Seoul 08826. South Korea

Background

In mice, neurogenesis is complete within 21 days after birth, yet adult brains retain their ability for neurogenesis in some specific areas, including the subgranular zone of the dentate gyrus and the subventricular zone of the lateral ventricle [1, 2]. These endogenous origins of neurogenesis seem to be an ideal source of compensatory repair and system functionality. For example, newborn cells in the subgranular zone of the dentate gyrus can migrate to the granular cell layer, where they integrate into the neuronal circuitry of the dentate gyrus as granule neurons [3]. These integrated neurons can then play a role in memory formation in the hippocampus. Adult neurogenesis can be affected by variables, such as environmental factors, growth factors, neurotransmitters, and external stimuli that alter the affective state of an animal [4-6]. Herbal supplementation can also modulate neurogenesis and thereby affect hippocampal functions, such as memory [7-9]. However, few studies have focused on the oil components of foods consumed in high quantities in oriental countries.

Vegetables of the genus Allium, such as garlic, onions, green onions, and chives, have been used as food additives in China and Egypt, and are now being studied for the prevention of infection [10-12] and cancer [13, 14]. The characteristic odor of garlic and chives arises from allicin (allyl 2-propene thiosulfinate or diallyl thiosulfinate) and other oil-soluble sulfur components, such as diallyl disulfide (DADS), diallyl trisulfide (DATS), and diallyl sulfide (DAS) [15-18]. These oil-soluble components exhibit antioxidant properties, by reducing the levels of reactive oxygen species and increasing glutathione-S-transferase expression [19, 20]. In previous studies, we demonstrated that S-allyl-L-cysteine, a watersoluble component of Allium species [21, 22], promotes cell proliferation and neuroblast differentiation in the dentate gyrus [7]. In addition, we have shown the neuroprotective effects of Z-ajoene, an oil-soluble component, against ischemic damage in the gerbil hippocampus [9]. On the other hand, the administration of 10 mg/kg DADS affects neurogenesis, by reducing hippocampal brain-derived neurotrophic factor (BDNF) levels, and impairs performance in the passive avoidance test [23]. In addition, the garlic essential oil (GO) compounds, DAS and DADS, promote the intrinsic calpain-caspase cascade for apoptosis in human neuroblastoma SH-SY5Y cells [24].

However, few studies have been conducted on the effects of oils from garlic or chives extracts on neurogenesis and their related mechanisms of action in the dentate gyrus. In the present study, we investigated the effects of GO and chives essential oil (CO) on cell proliferation and neuroblast differentiation in the naïve

mouse, and considered possible mechanisms responsible for these effects in the dentate gyrus.

Methods

Experimental animals

Male C57BL/6 mice were purchased from Japan SLC Inc. (Shizuoka, Japan). They were housed under standard conditions with temperature (22 °C) and humidity (60 %) control, a 12-h light/dark cycle, and free access to food and water. The handling and care of the animals conformed to the guidelines complying with current international laws and policies (NIH Guide for the Care and Use of Laboratory Animals, NIH Publication No. 85–23, 1985, revised 1996) and were approved by the Institutional Animal Care and Use Committee (IACUC) of Seoul National University (Approval number: SNU-141112-1). During all of the experiments, every effort was made to minimize the number of animals used and the suffering caused by the procedures employed in the present study.

Preparation of essential garlic and chives oils

Garlic (Allium sativum) and chives (Allium hookeri) (1 kg) were purchased from a local market in South Korea. They were authenticated by two oriental medicine doctors (Dr. Gwang Lim Choi and Cheol Soo Lee, Kyung-Dong Oriental Medical Clinic, Seoul South Korea) and the voucher specimen was deposited in our laboratory (deposition number: 2015-003). The oils can be obtained upon request via email to the corresponding authors. Both essential oils were obtained by steam distillation from Allium sativum and Allium hookeri. Briefly, the crushed garlic clove (200 g) or chives (200 g) was placed in a 2 L round-bottom flask and distilled water was added (400 mL) with boiling bubble stone. Then, the mixture was sonicated for 20 min. Finally, the mixture of garlic-water or chiveswater was distilled for 4 h at 100 °C using a Clevenger-type apparatus, according to the method of Dadalioglu and Evrendilek [25].

Treatment with essential garlic or chives oil

The animals were divided into 3 groups (n = 10 in each group): vehicle (corn oil)-, 10 ml/kg GO-, and 10 ml/kg CO-treated group. Vehicle, GO, or CO was administered orally to 9-week-old mice once a day for 3 weeks. The dosage was chosen, because oral administration of garlic oil (5 ml/kg body weight) daily for 3 months has been shown to significantly improve NaNO₂-induced neurobiochemical disorders and oxidative stress [26]. The schedule was adopted, because doublecortin (DCX) is exclusively expressed in immature neurons from 1 to 28 days of cell age [27, 28].

Novel object recognition test

The apparatus consisted of an acrylic box with three opaque walls and one transparent wall ($45 \times 45 \times 30$ cm). The floor was covered with woodchip bedding, which was mixed between trials and testing days to prevent the build-up of odor in any particular place. The test objects were made of solid metal and they could not be displaced by the mice because of their weight. The objects were cleaned with bleach to remove residual odors.

At 20 days of vehicle, GO, or CO treatment (1 h after dose was provided), each mouse (n = 10 per group) was allowed to explore the apparatus for 2 min. On the testing day (at 21 days of treatment), 1 h following the last dose, a session of two 2-min trials was performed. In the "sample" trial (T1), two identical objects were placed in two opposite corners of the apparatus. A mouse was placed in the apparatus and was left to explore these two identical objects. After T1, the mouse was placed back in its home cage for an inter-trial interval of 1 h. Subsequently, a "choice" trial (T2) was performed. In T2, a new object (N) replaced one of the objects presented in T1. The mice were exposed to two different objects: the familiar (F) and the new one (N). Exploration was defined as directing the nose towards the object at a distance of no more than 2 cm and/or touching the object with the nose. From this measure a series of variables were then calculated: the total time spent exploring the two identical objects in T1 and the time spent exploring the two different objects, F and N, in T2.

The preference for F and N in T2 was determined, by comparing the time spent exploring F with that spent exploring N. The discrimination index (DI) represents the difference in exploration time expressed as a proportion of the total time spent exploring the two objects in T2.

Tissue processing

For histology, the animals (n=5 in each group) from the vehicle-, GO-, or CO-treated groups were anesthetized with 1 g/kg urethane (Sigma-Aldrich, St. Louis, MO, USA) 2 h after novel object recognition test and perfused transcardially with 0.1 M of phosphate-buffered saline (PBS, pH 7.4) followed by 4 % paraformaldehyde in 0.1 M phosphate-buffer (pH 7.4). Brains were removed and post-fixed in a fixative for 12 h. Brain tissues were cryoprotected with 30 % sucrose overnight. Brain sections of 30- μ m thickness were serially cut in the coronal plane using a cryostat (Leica, Wetzlar, Germany). Sections were collected in six-well plates containing PBS for further processing.

Immunohistochemistry

In order to obtain accurate data for immunohistochemistry, free-floating sections were carefully processed

under the same conditions. For each animal, tissue sections were selected between 1.46 mm and 2.46 mm posterior to bregma, by referring to the mouse atlas [29]. Ten sections, 90 µm apart from each other, were obtained and sequentially treated with 0.3 % hydrogen peroxide in 0.05 M PBS and 10 % horse serum in 0.05 M PBS. They were then incubated overnight with diluted rabbit anti-Ki67 antibody (1:1,000; Abcam, Cambridge, UK) or goat anti-DCX antibody (1:50; Santa Cruz Biotechnology, Santa Cruz, CA, USA), and subsequently exposed to biotinylated rabbit anti-goat or goat anti-rabbit IgG (diluted 1:200; Vector, Burlingame, CA, USA) and streptavidin peroxidase complex (diluted 1:200, Vector). Then, the sections were visualized by reaction with 3,3'-diaminobenzidine tetrahydrochloride (Sigma-Aldrich).

Number of Ki67- and DCX-positive cells was counted in each section of the dentate gyrus by using an image analysis system equipped with a computer-based CCD camera (software: Optimas 6.5, CyberMetrics, Scottsdale, AZ, USA). Cell counts from all of the sections of all of the mice in each group were averaged.

BDNF protein levels in the hippocampal homogenates

For BDNF protein level and acetylcholinesterase (AChE) activity analyses, the animals (n = 5 in each group) from the vehicle-, GO-, or CO-treated groups were anesthetized with 2 g/kg urethane (Sigma-Aldrich) at 2 h after novel object recognition test and the hippocampi were dissected from the brain tissue. Left and right side of the hippocampus was used for BDNF protein levels and AChE activity assessment, respectively. BDNF protein level in the left part of the hippocampus was measured using a BDNF Emax immunoassay kit (Promega, Madison, WI, USA), as described previously [30]. Briefly, the tissue samples were weighed and 300 µL of lysis buffer was added to each sample. The samples were then sonicated for 30 s and centrifuged at 4 °C for 20 min. The supernatant was stored at -20 °C until it was analyzed. All samples were assayed in duplicate and the absorbance was read with an enzymelinked immunosorbent assay (ELISA) plate reader (BioTek, Winooski, VT, USA). Total protein concentrations were estimated using BioRAD procedure (Hercules, CA, USA). The concentration of each sample was calculated by plotting the absorbance values on a standard curve generated by the assay.

Measurement of AChE activity in the hippocampus

For measuring of the AChE activity in the synaptosome, the right part of hippocampus was homogenized in 10 volumes of an ice-cold medium, consisting of 320 mM sucrose, 0.1 mM EDTA, and 5 mM 4-(2-hydroxyethyl)1-piperazineethanesulfonic acid (pH 7.5) in a motor

driven Teflon-glass homogenizer. The synaptosomes were isolated, as described previously [31], using a discontinuous Percoll gradient. The pellet was suspended in an isoosmotic solution and the final protein concentration was adjusted to 0.5 mg/mL. Synaptosomes were prepared fresh daily, maintained at 4 °C throughout the procedure and used for enzymatic assays. The AChE enzymatic assay outcome was determined with a modified spectrophotometric method [32, 33].

The reaction mixture (2 mL final volume) contained 100 mM K⁺-phosphate buffer (pH 7.5) and 1 mM 5,5′-dithio-bis-(2-nitrobenzoic) acid. The catalytic activity was measured based on AChE reaction with 5,5′-dithio-bis-(2-nitrobenzoic acid) during 2-min incubation period at 25 °C, by the measurement of absorbance at 412 nm of the yellow anion, 2-nitro-5-thio-benzoate, produced from thiocholine. The enzyme was pre-incubated for 2 min. The reaction was initiated by adding 0.8 mM acetylthiocholine iodide (ATCh). All samples were run in triplicate and enzyme activity was expressed in μ mol ATCh·h⁻¹/mg of protein.

Statistical analysis

Data are expressed as means \pm standard error of the mean (SEM). Differences among the means were statistically

analyzed with a one-way analysis of variance, followed by Bonferroni's post-hoc test, in order to compare the effects of GO and CO on: novel object recognition, cell proliferation, neuroblast differentiation, BDNF protein levels, and AChE activity in mice, using GraphPad Prism 5.01 software (GraphPad Software, Inc., La Jolla, CA, USA). The statistical significance level was set at P < 0.05.

Results

Effects of GO or CO on novel object recognition

During the training period, all mice from the vehicle-, GO-, or CO-treated groups spent the same amount of time exploring the two objects; however, during the test period, mice spent more time exploring the novel object than the familiar one. However, the proportions of time were different between the vehicle-, GO-, and CO-treated groups. The mice from the GO-treated group spent significantly more time exploring the novel object than mice in the vehicle-treated group. In contrast, mice in the CO-treated group spent significantly less time exploring the novel object than mice in the vehicle-treated group (Fig. 1). DI values significantly increased in the GO-treated group compared to that in the vehicle-treated group, while they significantly decreased in the CO-treated group (Fig. 1).

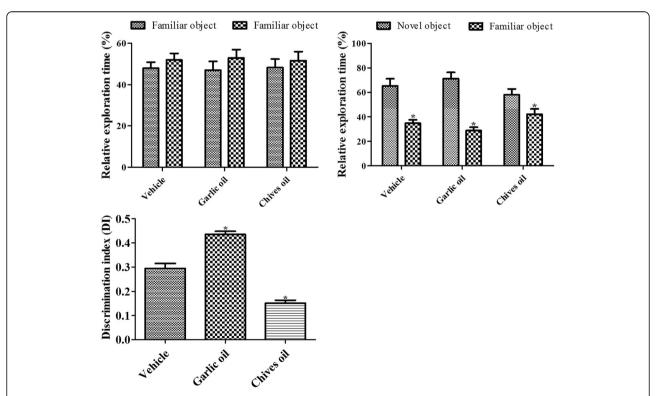


Fig. 1 The effect of vehicle, garlic essential oil and chives essential oil on exploration time and discrimination index of familiar vs. novel objects in the novel object recognition test in mice (n = 10 per group; *p < 0.05 vs. familiar object on exploration time or vs. vehicle-treated group in discrimination index). Data for exploration time for each object (one object was replaced by a new one on the testing day) are presented as a percentage of total exploration time. All data are shown as mean exploration time \pm SEM

Effects of GO or CO on cell proliferation

In the vehicle-treated group, Ki67-positive nuclei were found in the subgranular zone of the dentate gyrus (Fig. 2a) and the average number of Ki67-immunoreactive nuclei was 9.1 per section (Fig. 2d). In the subgranular zone of the dentate gyrus of the GO-treated group, Ki67-positive nuclei were more abundant in comparison to those in the in vehicle-treated group, and the average number of the nuclei was 14.7 per section (Fig. 2b and d). In the CO-treated group, few Ki67-positive nuclei were detected in the subgranular zone of the dentate gyrus and there were 3.8 positive nuclei per section (Fig. 2c and d).

Effects of GO or CO on neuroblast differentiation

In the vehicle-treated group, DCX-immunoreactive neuroblasts were detected in the subgranular zone of the dentate gyrus and their dendrites extended into the molecular layer of the dentate gyrus (Fig. 3a and b). In this group, the average number of DCX-immunoreactive neuroblasts was 75.4 per section (Fig. 3g). In the GO-treated group, DCX-immunoreactive neuroblasts were more abundant in the subgranular zone of the dentate gyrus compared to those of the vehicle-treated group (Fig. 3c and d) and the average number of DCX-immunoreactive neuroblasts was 126.4 per section (Fig. 3g). In the CO-treated group, only a few DCX-

immunoreactive neuroblasts were detected in the subgranular zone of the dentate gyrus and their dendrites were poorly detected (Fig. 3e and f). In this group, the average number of DCX-immunoreactive neuroblasts was 41.5 per section (Fig. 3g).

Effects of GO or CO on BDNF protein levels

BDNF protein levels significantly increased in the dentate gyrus homogenates of the GO-treated group compared to those in the vehicle-treated group. In contrast, BDNF protein levels significantly decreased in the CO-treated group compared to those in the vehicle-treated group (Fig. 4).

Effects of GO or CO on AChE activity

In the vehicle-treated group, AChE activity was 0.112 μ mol ATCh-h⁻¹/mg protein in the hippocampal homogenates. In the GO-treated group, AChE activity in the synaptosome was significantly decreased, representing only 71.3 % of the activity of the vehicle-treated group. However, in the CO-treated group, AChE activity was similar to that in the vehicle-treated group (Fig. 4).

Discussion

Allium species, such as garlic and chives, contain various organosulfur compounds, such as ajoene, vinyldithiins,

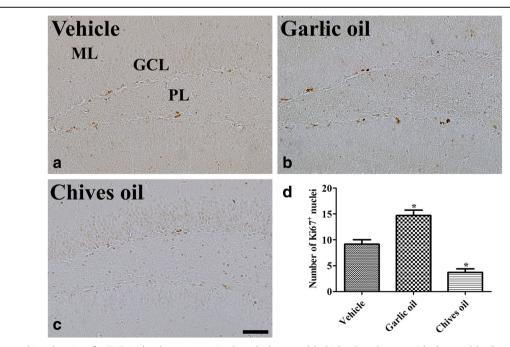


Fig. 2 Immunohistochemistry for Ki67 in the dentate gyrus in the vehicle-treated (vehicle, **a**), garlic essential oil-treated (garlic oil, **b**) and chives essential oil-treated (chives oil, **c**) groups. Ki67-positive ($^+$) nuclei are observed in the subgranular zone of dentate gyrus. Note that Ki67 $^+$ nuclei are abundant in the garlic essential oil-treated group, while in the chives essential oil-treated group, Ki67 $^+$ nuclei are few. GCL, granule cell layer; ML, molecular layer; PL, polymorphic layer. Scale bar = 50 μm. **d** Number of Ki67 $^+$ nuclei per section for each group (n = 5 per group; *p < 0.05, versus vehicle group). Data are presented as mean ± SEM

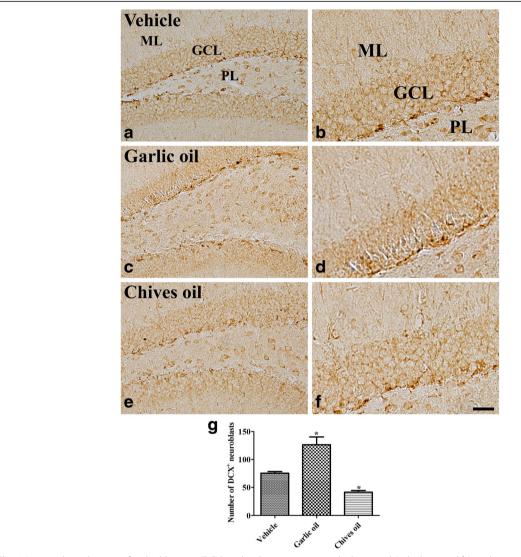


Fig. 3 Immunohistochemistry for doublecortin (DCX) in the dentate gyrus in vehicle-treated (vehicle, **a**, and **b**), garlic essential oil-treated (garlic oil, **c**, and **d**) and chives essential oil-treated (chives oil, **e**, and **f**) groups. DCX-immunoreactive (†) neuroblasts are seen in the subgranular zone of the dentate gyrus and their dendrites are observed in the molecular layer (ML) of the dentate gyrus. Note that DCX⁺ neuroblasts and their dendrites are abundant in the garlic essential oil-treated group, while in the chives essential oil-treated group, DCX⁺ neuroblasts and their dendrites are few. GCL, granule cell layer; ML, molecular layer; PL, polymorphic layer. Scale bar = $50 \mu m$ (**a**, **c**, **e**), $25 \mu m$ (**b**, **d**, **f**). **g** Number of DCX⁺ neuroblasts per section for each group ($n = 5 \mu m$ group; *p < 0.05, versus vehicle group). Data are presented as mean \pm SEM

DADS, and DATS [34–38]. In the present study, we focused on the effects of GO and CO on the hippocampus-dependent neurogenesis and memory formation in naïve mice. Significant improvements in the novel object recognition were observed in the GO-treated group, while the CO-treated group showed a significant reduction of the fraction of time spent exploring the novel object compared to that of the vehicle-treated group. These results are consistent with those of previous studies, which showed that sprouted or crude garlic extracts improved scopolamine-induced impairments of memory and cognition in mice [39]. It has also been

reported that repeated administration of aged garlic extract enhanced memory function by increasing 5-hydroxytryptamine levels in rats [40]. Furthermore, commercial garlic extract powder capsules have been shown to improve cognitive function and brain mitochondrial function, which were impaired in obese, insulin-resistant rats because of high-fat diet [41].

The hippocampus is a major region related to memory formation and cognition, with continuous neurogenesis occurring in adult life [42–44]. It has been reported that enhanced hippocampal cell proliferation and neurogenesis improves memory deficits and memory impairments

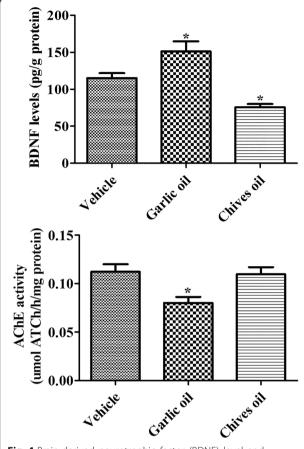


Fig. 4 Brain-derived neurotrophic factor (BDNF) level and acetylcholinesterase (AChE) activity in synaptosomes of the hippocampi of vehicle-treated (vehicle), garlic essential oil-treated (garlic oil) and chives essential oil-treated (chives oil) groups (n = 5 per group; *p < 0.05, versus vehicle group). Data are presented as mean \pm SEM

[45-48]. In this study, we observed significant increases in the number of cells positive for Ki67 and DCX, which are markers for proliferating cells and neuroblasts, respectively, in the subgranular zone of the dentate gyrus in the GO-treated group. However, in the CO-treated group, proliferation and differentiation significantly reduced compared to that of the control group. Previous studies have reported that garlic extracts and ascorbic acid ameliorated lead-induced neurotoxicity and decreased the number of DCX-positive neuroblasts [49]. These results suggest that increased cell proliferation and differentiation into DCX-positive neuroblasts by GO treatment may be related to the improvement of novel object recognition in the intact hippocampus. However, CO treatment may reduce novel object recognition in the hippocampus of naïve mice.

To identify possible mechanisms responsible for the enhancement of neurogenesis by GO in this study, we examined changes in the BDNF levels in the hippocampus.

BDNF is implicated, as a potent neurotrophic factor regulating adult neurogenesis as well as synaptic transmission in the brain [50-55]. In addition, acetylcholine (ACh) and AChE activity are related to hippocampal neurogenesis [56-58]. Overexpression of the vesicular ACh transporter has been reported to enhance dendritic complexity in adult-born hippocampal neurons [59]. In addition, ACh enhanced cell proliferation and DCX-positive neuroblast production from neural stem cells in vitro [57, 60]. Furthermore, AChE inhibitors, such as donepezil hydrochloride, have been shown to enhance neurogenesis via downregulation of AChE activity in mice with vascular dementia [61, 62]. Finally, we have previously demonstrated that inhibition of AChE or butyrylcholinesterase activity significantly increases the cell proliferation and neuroblast differentiation in the dentate gyrus [63].

In the present study, we observed significantly increased BDNF levels in the hippocampus of the GO-treated group, but not in the CO-treated group, when compared to the control group. AChE activity was significantly decreased in the GO-treated group, but similarly detected in the CO-treated group, with the AChE activity inversely proportional to the BDNF levels in GO-treated group. Many previous studies have reported that BDNF may confer protective effects against various neurotoxic conditions in the brain, via reduction of the AChE levels or activity [64-66]. In addition, emerging evidence suggests that regulation of AChE activity by BDNF is related to hippocampal neurogenesis [61, 67-69]. For example, huperzine A, an AChE inhibitor, increases BDNF mRNA and protein levels, while cholinergic denervation or muscarinic antagonist treatment, such as atropine, decreases the hippocampal BDNF mRNA levels [70]. Interestingly, while garlic and chives are both representatives of the Allium species, their essential oils showed contrasting effects on novel object cognition, cell proliferation, neuroblast differentiation, BDNF levels, and AChE activity. In fact, it has been reported that various effects on neuroprotection, memory, and cell proliferation in the brain result from compounds of different Allium species. For example, Z-ajoene from garlic ameliorates the scopolamine-induced memory impairment dosedependently [71]; however, allin and DADS do not improve memory performance, cell proliferation, or neuroblast differentiation in the same model [23, 71, 72]. Therefore, the contradictory results of this study may be explained by the differing composition ratios and by how these compounds interact with memory formation in the naïve mouse. The exact mechanisms by which components induce the different effects of garlic and chives remain to be elucidated. In the present study, we chose one dosage of GO and CO. Test with varying doses would be needed for human trials, to use GO as functional food.

In addition, the comparative chemical analytic studies of GO and CO will help detect possible toxic compounds in these *Allium* species.

Conclusion

In conclusion, GO could be helpful to promote cell proliferation, neuroblast differentiation, novel object recognition, and memory formation by modulating BDNF levels and AChE activity in the hippocampus.

Abbreviations

ACh: Acetylcholine; AChE: Acetylcholinesterase; ATCh: Acetylthiocholine iodide; BDNF: Brain-derived neurotrophic factor; DADS: Diallyl disulfide; DAS: Diallyl sulfide; DATS: Diallyl trisulfide; DCX: Doublecortin; SEM: Standard error of mean

Funding

This research was supported by Korea Food Research Institute (E0143023852) and by 2014 Research Grant from Kangwon National University. This study was partially supported by the Research Institute for Veterinary Science, Seoul National University.

Availability of data and materials

The supporting materials including garlic and chives oil can be obtained upon request via email to the corresponding authors.

Authors' contributions

All authors conceived the study. HYJ, KYL, JHC and IKH designed the study. HYJ and KYL wrote the manuscript and JHC and IKH edited the manuscript. HYJ, DYY and JWK conducted the animal experiments. KYL and JHC measured brain-derived neurotrophic factors and acetylcholinesterase activity. MY, SL, KYY participated in designing and discussing the study. All authors have read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Consent for publication

This information is not relevant.

Ethics approval and consent to participate

The handling and care of the animals conformed to the guidelines established to comply with current international laws and policies (NIH Guide for the Care and Use of Laboratory Animals, NIH Publication No. 85–23, 1985, revised 1996) and were approved by the Institutional Animal Care and Use Committee (IACUC) of Seoul National University (Approval number: SNU-141112-1).

Author details

¹Department of Anatomy and Cell Biology, College of Veterinary Medicine, Research Institute for Veterinary Science, Seoul National University, Seoul 08826, South Korea. ²Department of Anatomy, College of Veterinary Medicine and Institute of Veterinary Science, Kangwon National University, Chuncheon 24341, South Korea. ³Food Analysis Center, Korea Food Research Institute, Sungnam 13539, South Korea. ⁴Department of Oral Anatomy, Research Institute of Oral Sciences, College of Dentistry, Gangneung-Wonju National University, Gangneung 25457, South Korea.

Received: 21 June 2016 Accepted: 6 October 2016 Published online: 03 November 2016

References

- Gage FH, Kempermann G, Palmer TD, Peterson DA, Ray J. Multipotent progenitor cells in the adult dentate gyrus. J Neurobiol. 1998;36:249–66.
- Gould E, Tanapat P, Hastings NB, Shors TJ. Neurogenesis in adulthood: a possible role in learning. Trends Cogn Sci. 1999;3:186–92.
- Jin K, Minami M, Lan JQ, Mao XO, Batteur S, Simon RP, Greenberg DA. Neurogenesis in dentate subgranular zone and rostral subventricular zone after focal cerebral ischemia in the rat. Proc Natl Acad Sci U S A. 2001;98:4710–5.

- LaDage LD. Environmental change, the stress response, and neurogenesis. Integr Comp Biol. 2015;55:372–83.
- Mendez-David I, Hen R, Gardier AM, David DJ. Adult hippocampal neurogenesis: an actor in the antidepressant-like action. Ann Pharm Fr. 2013;71:143–9.
- Sequerra EB, Costa MR, Menezes JR, Hedin-Pereira C. Adult neural stem cells: plastic or restricted neuronal fates? Development. 2013;140:3303–9.
- Nam SM, Yoo DY, Kim W, Yoo M, Kim DW, Won MH, Hwang IK, Yoon YS. Effects of S-allyl-L-cysteine on cell proliferation and neuroblast differentiation in the mouse dentate gyrus. J Vet Med Sci. 2011;73:1071–5.
- Ye M, Chung HS, An YH, Lim SJ, Choi W, Yu AR, Kim JS, Kang M, Cho S, Shim I, Bae H. Standardized herbal formula PM012 decreases cognitive impairment and promotes neurogenesis in the 3xTg AD mouse model of Alzheimer's disease. Mol Neurobiol. doi: 10.1007/s12035-015-9458-x.
- Yoo DY, Choi JH, Kim W, Jung HY, Nam SM, Kim JW, Yoon YS, Yoo KY, Won MH, Hwang IK. Cynomorium songaricum extract enhances novel object recognition, cell proliferation and neuroblast differentiation in the mice via improving hippocampal environment. BMC Complement Altern Med. 2014;14:5.
- Arreola R, Quintero-Fabián S, López-Roa RI, Flores-Gutiérrez EO, Reyes-Grajeda JP, Carrera-Quintanar L, Ortuño-Sahagún D. Immunomodulation and anti-inflammatory effects of garlic compounds. J Immunol Res. 2015;2015;401630.
- Kyung KH. Antimicrobial properties of allium species. Curr Opin Biotechnol. 2012;23:142–7.
- Lazarević JS, Ethordevic AS, Zlatkovic BK, Radulovic NS, Palic RM. Chemical composition and antioxidant and antimicrobial activities of essential oil of Allium sphaerocephalon L. subsp. sphaerocephalon (Liliaceae) inflorescences. J Sci Food Agric. 2011;91:322–9.
- 13. Lee SK, Park YJ, Ko MJ, Wang Z, Lee HY, Choi YW, Bae YS. A novel natural compound from garlic (*Allium sativum* L.) with therapeutic effects against experimental polymicrobial sepsis. Biochem Biophys Res Commun. 2015;464:774–9.
- Wang HC, Yang JH, Hsieh SC, Sheen LY. Allyl sulfides inhibit cell growth of skin cancer cells through induction of DNA damage mediated G2/M arrest and apoptosis. J Agric Food Chem. 2010;58:7096–103.
- Amagase H, Petesch BL, Matsuura H, Kasuga S, Itakura Y. Intake of garlic and its bioactive components. J Nutr. 2001;131:955S–62
- Craig WJ. Health-promoting properties of common herbs. Am J Clin Nutr. 1999;70(3 Suppl):4915–9.
- Kim NY, Park MH, Jang EY, Lee J. Volatile distribution in garlic (Allium sativum L.) by solid phase microextraction (SPME) with different processing conditions. Food Sci Biotechnol. 2011;20:775–82.
- Li R, Chen WC, Wang WP, Tian WY, Zhang XG. Extraction of essential oils from garlic (*Allium sativum*) using ligarine as solvent and its immunity activity in gastric cancer rat. Med Chem Res. 2009;19:1092–105.
- Iciek MB, Kowalczyk-Pachel D, Kwiecien I, Dudek MB. Effects of different garlic-derived allyl sulfides on peroxidative processes and anaerobic sulfur metabolism in mouse liver. Phytother Res. 2012;26:425–31.
- Maldonado PD, Alvarez-Idaboy JR, Aguilar-González A, Lira-Rocha A, Jung-Cook H, Medina-Campos ON, Pedraza-Chaverrí J, Galano A. Role of allyl group in the hydroxyl and peroxyl radical scavenging activity of Sallylcysteine. J Phys Chem B. 2011;115:13408–17.
- 21. Borek C. Antioxidant health effects of aged garlic extract. J Nutr. 2001;131:10105–5.
- Brunetti L, Menghini L, Orlando G, Recinella L, Leone S, Epifano F, Lazzarin F, Chiavaroli A, Ferrante C, Vacca M. Antioxidant effects of garlic in young and aged rat brain in vitro. J Med Food. 2009;12:1166–9.
- 23. Ji ST, Kim MS, Park HR, Lee E, Lee Y, Jang YJ, Kim HS, Lee J. Diallyl disulfide impairs hippocampal neurogenesis in the young adult brain. Toxicol Lett. 2013;221:31–8.
- Karmakar S, Banik NL, Patel SJ, Ray SK. Garlic compounds induced calpain and intrinsic caspase cascade for apoptosis in human malignant neuroblastoma SH-SY5Y cells. Apoptosis. 2007;12:671–84.
- Dadalioglu I, Evrendilek GA. Chemical compositions and antibacterial effects
 of essential oils of Turkish oregano (*Origanum minutiflorum*), bay laurel
 (*Laurus nobilis*), Spanish lavender (*Lavandula stoechas* L.), and fennel
 (*Foeniculum vulgare*) on common foodborne pathogens. J Agric Food
 Chem. 2004;52:8255–60.
- Hassan HA, Hafez HS, Zeghebar FE. Garlic oil as a modulating agent for oxidative stress and neurotoxicity induced by sodium nitrite in male albino rats. Food Chem Toxicol. 2010;48:1980–5.

- Brown JP, Couillard-Despres S, Cooper-Kuhn CM, Winkler J, Aigner L, Kuhn HG. Transient expression of doublecortin during adult neurogenesis. J Comp Neurol. 2003;467:1–10.
- Couillard-Despres S, Winner B, Schaubeck S, Aigner R, Vroemen M, Weidner N, Bogdahn U, Winkler J, Kuhn HG, Aigner L. Doublecortin expression levels in adult brain reflect neurogenesis. Eur J Neurosci. 2005;21:1–14.
- Franklin KBJ, Paxinos G. The mouse brain in stereotaxic coordinates.
 San Diego: Academic; 1997.
- Yoo DY, Kim W, Nam SM, Yoo KY, Lee CH, Choi JH, Won MH, Hwang IK, Yoon YS. Reduced cell proliferation and neuroblast differentiation in the dentate gyrus of high fat diet-fed mice are ameliorated by metformin and glimepiride treatment. Neurochem Res. 2011;36:2401–8.
- Nagy A, Delgado-Escueta AV. Rapid preparation of synaptosomes from mammalian brain using nontoxic isoosmotic gradient material (Percoll).
 J Neurochem. 1984;43:1114–23.
- Ellman GL, Courtney KD, Andres Jr V, Feather-Stone RM. A new and rapid colorimetric determination of acetylcholinesterase activity. Biochem Pharmacol. 1961;7:88–95.
- Rocha JB, Emanuelli T, Pereira ME. Effects of early undernutrition on kinetic parameters of brain acetylcholinesterase from adult rats. Acta Neurobiol Exp (Wars), 1993:53:431–7.
- Foskolos A, Siurana A, Rodriquez-Prado M, Ferret A, Bravo D, Calsamiglia S.
 The effects of a garlic oil chemical compound, propyl-propane
 thiosulfonate, on ruminal fermentation and fatty acid outflow in a dualflow continuous culture system. J Dairy Sci. 2015;98:5482–91.
- Ito Y, Kosuge Y, Sakikubo T, Horie K, Ishikawa N, Obokata N, Yokoyama E, Yamashina K, Yamamoto M, Saito H, Arakawa M, Ishige K. Protective effect of S-allyl-L-cysteine, a garlic compound, on amyloid β-protein-induced cell death in nerve growth factor-differentiated PC12 cells. Neurosci Res. 2003;46:119–25.
- 36. Staba EJ, Lash L, Staba JE. A commentary on the effects of garlic extraction and formulation on product composition. J Nutr. 2001;131:1118S–9.
- Ferri N, Yokoyama K, Sadilek M, Paoletti R, Apitz-Castro R, Gelb MH, Corsini A. Ajoene, a garlic compound, inhibits protein prenylation and arterial smooth muscle cell proliferation. Br J Pharmacol. 2003;138:811–8.
- Ivanova A, Mikhova B, Najdenski H, Tsvetkova I, Kostova I. Chemical composition and antimicrobial activity of wild garlic Allium ursinum of Bulgarian origin. Nat Prod Commun. 2009;4:1059–62.
- Mukherjee D, Banerjee S. Learning and memory promoting effects of crude garlic extract. Indian J Exp Biol. 2013;51:1094–100.
- Haider S, Naz N, Khaliq S, Perveen T, Haleem DJ. Repeated administration of fresh garlic increases memory retention in rats. J Med Food. 2008;11:675–9.
- Pintana H, Sripetchwandee J, Supakul L, Apaijai N, Chattipakorn N, Chattipakorn S. Garlic extract attenuates brain mitochondrial dysfunction and cognitive deficit in obese-insulin resistant rats. Appl Physiol Nutr Metab. 2014;39:1373–9.
- 42. Epp JR, Chow C, Galea LA. Hippocampus-dependent learning influences hippocampal neurogenesis. Front Neurosci. 2013;7:57.
- 43. Frankland PW, Josselyn SA. Hippocampal neurogenesis and memory clearance. Neuropsychopharmacology. 2016;41:382–3.
- Kempermann G, Song H, Gage FH. Neurogenesis in the adult hippocampus. Cold Spring Harb Perspect Med. 2015;5:a018812.
- 45. Cameron HA, Glover LR. Adult neurogenesis: beyond learning and memory. Annu Rev Psychol. 2015;66:53–81.
- Gundersen BB, Briand LA, Onksen JL, Lelay J, Kaestner KH, Blendy JA. Increased hippocampal neurogenesis and accelerated response to antidepressants in mice with specific deletion of CREB in the hippocampus: role of cAMP response-element modulator tau. J Neurosci. 2013;33:13673–85.
- Kesner RP, Xu H, Sommer T, Wright C, Barrera VR, Fanselow MS. The role of postnatal neurogenesis in supporting remote memory and spatial metric processing. Hippocampus. 2015;25:1072.
- Lilja AM, Malmsten L, Röjdner J, Voytenko L, Verkhratsky A, Ögren SO, Nordberg A, Marutle A. Neural stem cell transplant-induced effect on neurogenesis and cognition in Alzheimer Tg2576 mice is inhibited by concomitant treatment with amyloid-lowering or cholinergic α7 nicotinic receptor drugs. Neural Plast. 2015;2015:370432.
- Alipour F, Bideskan AE, Fazel A, Sadeghi A, Hami J, Kheradmand H, Haghir H. Protective effects of ascorbic acid and garlic extract against neurogenesis inhibition caused by developmental lead exposure in the dentate gyrus of rat. Comp Clin Pathol. 2014;23:1681–7.

- Bath KG, Akins MR, Lee FS. BDNF control of adult SVZ neurogenesis. Dev Psychobiol. 2012;54:578–89.
- Botterill JJ, Brymer KJ, Caruncho HJ, Kalynchuk LE. Aberrant hippocampal neurogenesis after limbic kindling: relationship to BDNF and hippocampaldependent memory. Epilepsy Behav. 2015;47:83–92.
- Le Hellard S, Håvik B, Espeseth T, Breilid H, Løvlie R, Luciano M, Gow AJ, Harris SE, Starr JM, Wibrand K, Lundervold AJ, Porteous DJ, Bramham CR, Deary IJ, Reinvang I, Steen VM. Variants in doublecortin- and calmodulin kinase like 1, a gene up-regulated by BDNF, are associated with memory and general cognitive abilities. PLoS One. 2009;4:e7534.
- Quesseveur G, David DJ, Gaillard MC, Pla P, Wu MV, Nguyen HT, Nicolas V, Auregan G, David I, Dranovsky A, Hantraye P, Hen R, Gardier AM, Déglon N, Guiard BP. BDNF overexpression in mouse hippocampal astrocytes promotes local neurogenesis and elicits anxiolytic-like activities. Transl Psychiatry. 2013;3:e253.
- Rossi C, Angelucci A, Costantin L, Braschi C, Mazzantini M, Babbini F, Fabbri ME, Tessarollo L, Maffei L, Berardi N, Caleo M. Brain-derived neurotrophic factor (BDNF) is required for the enhancement of hippocampal neurogenesis following environmental enrichment. Eur J Neurosci. 2006;24:1850–6.
- Wei Z, Liao J, Qi F, Meng Z, Pan S. Evidence for the contribution of BDNF-TrkB signal strength in neurogenesis: an organotypic study. Neurosci Lett. 2015;606:48–52.
- Ijomone OM, Nwoha PU. Nicotine inhibits hippocampal and striatal acetylcholinesterase activities, and demonstrates dual action on adult neuronal proliferation and maturation. Pathophysiology. 2015;22:231–9.
- Mohapel P, Leanza G, Kokaia M, Lindvall O. Forebrain acetylcholine regulates adult hippocampal neurogenesis and learning. Neurobiol Aging. 2005;26:939–46.
- Veena J, Rao BS, Srikumar BN. Regulation of adult neurogenesis in the hippocampus by stress, acetylcholine and dopamine. J Nat Sci Biol Med. 2011;2:26–37.
- Nagy PM, Aubert I. Overexpression of the vesicular acetylcholine transporter enhances dendritic complexity of adult-born hippocampal neurons and improves acquisition of spatial memory during aging. Neurobiol Aging. 2015;36:1881–9.
- Paez-Gonzalez P, Asrican B, Rodriguez E, Kuo CT. Identification of distinct ChAT⁺ neurons and activity-dependent control of postnatal SVZ neurogenesis. Nat Neurosci. 2014;17:934–42.
- Kotani S, Yamauchi T, Teramoto T, Ogura H. Donepezil, an acetylcholinesterase inhibitor, enhances adult hippocampal neurogenesis. Chem Biol Interact. 2008;175:227–30.
- Kwon KJ, Kim MK, Lee EJ, Kim JN, Choi BR, Kim SY, Cho KS, Han JS, Kim HY, Shin CY, Han SH. Effects of donepezil, an acetylcholinesterase inhibitor, on neurogenesis in a rat model of vascular dementia. J Neurol Sci. 2014:347:66–77.
- Yoo DY, Woo YJ, Kim W, Nam SM, Lee BH, Yeun GH, Yoon YS, Won MH, Park JH, Hwang IK. Effects of a new synthetic butyrylcholinesterase inhibitor, HBU-39, on cell proliferation and neuroblast differentiation in the hippocampal dentate gyrus in a scopolamine-induced amnesia animal model. Neurochem Int. 2011;59:722–8.
- Comim CM, Reis PA, Frutuoso VS, Fries GR, Fraga DB, Kapczinski F, Zugno AI, Barichello T, Quevedo J, Castro-Faria-Neto HC. Effects of experimental cerebral malaria in memory, brain-derived neurotrophic factor and acetylcholinesterase activity in the hippocampus of survivor mice. Neurosci Lett. 2012;523:104–7.
- Pandareesh MD, Anand T. Neuromodulatory propensity of Bacopa monniera against scopolamine-induced cytotoxicity in PC12 cells via down-regulation of AChE and up-regulation of BDNF and muscarnic-1 receptor expression. Cell Mol Neurobiol. 2013;33:875–84.
- Scherer EB, da Cunha MJ, Matte C, Schmitz F, Netto CA, Wyse AT.
 Methylphenidate affects memory, brain-derived neurotrophic factor
 immunocontent and brain acetylcholinesterase activity in the rat. Neurobiol
 Learn Mem. 2010;94:247–53.
- Bendix I, Serdar M, Herz J, von Haefen C, Nasser F, Rohrer B, Endesfelder S, Felderhoff-Mueser U, Spies CD, Sifringer M. Inhibition of acetylcholinesterase modulates NMDA receptor antagonist mediated alterations in the developing brain. Int J Mol Sci. 2014;15:3784–98.
- Lee JS, Kim HG, Lee HW, Han JM, Lee SK, Kim DW, Saravanakumar A, Son CG. Hippocampal memory enhancing activity of pine needle extract against scopolamine-induced amnesia in a mouse model. Sci Rep. 2015;5:9651.

- 69. Yoo DY, Choi JH, Kim W, Nam SM, Jung HY, Kim JH, Won MH, Yoon YS, Hwang IK. Effects of luteolin on spatial memory, cell proliferation, and neuroblast differentiation in the hippocampal dentate gyrus in a scopolamine-induced amnesia model. Neurol Res. 2013;35:813–20.
- Lapchak PA, Araujo DM, Hefti F. Cholinergic regulation of hippocampal brain-derived neurotrophic factor mRNA expression: evidence from lesion and chronic cholinergic drug treatment studies. Neuroscience. 1993;52:575–85.
- 71. Yamada N, Hattori A, Hayashi T, Nishikawa T, Fukuda H, Fujino T. Improvement of scopolamine-induced memory impairment by *Z*-ajoene in the water maze in mice. Pharmacol Biochem Behav. 2004;78:787–91.
- Miller Jr RR, Slathar JR, Luvisotto ML. Alpha-tocopherol and gammatocopherol attenuate ethanol-induced changes in membrane fatty acid composition in embryonic chick brains. Teratology. 2000;62:26–35.

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at www.biomedcentral.com/submit

